Vol. 1, No. 1, 44-52, 2016

## Antimicrobial Activity of Mangrove Actinomycetes from Soil Sample of Rhizophora Apiculata



e Science Publishing

T. Janaki<sup>1</sup>

<sup>1</sup>Department of Botany, K. M. Centre for P. G. Studies (Autonomous), Puducherry-8, India

### ABSTRACT

Totally 22 actinomycetes were isolated by dry heat (70oC) pre-treatment method on Starch casein agar media, from the soil sample that was collected nearer to the root region of the mangrove Rhizophora apiculata (Blume)-Rhizophoraceae from the back water area, Ariyankuppam, Puducherry (UT). All the 22 actinomycetes were subjected for primary screening against the 10 gram negative bacteria, 2 gram positive bacteria by agar plug method. The total percentage of inhibition by actinomycetes against bacteria in primary screening was noted as E.coli-9%, K. pneumoniae-0%, P.vulgaris-0%, P.aeruginosa-31.8%, S.typhi-13.6%, S.flexneri-13.6%, V.cholera-9%, B.bronchiseptica-68.2%, P.fluorescens-0%, E.faecalis-13.6%, B.subtilis-40.9%, S.aureus-9%. Totally 20 (90.91%) actinomycetes showed antibacterial activity towards any one of the tested bacteria, 2 (9.09%) actinomycetes showed no antagonistic activity. From these, it was noted that the mangrove actinomycetes were strong in their antibacterial activity. Only 4 actinomycetes were selected from R. apiculata and that were subjected for secondary screening. Out of 22 isolates from R.apiculata, 8 (36.36%) were active and 14 (63.64%) were inactive for the Candida albicans. 3 isolates from R.apiculata with strong anticandida activity were selected and subjected to confirmatory screening using cross streak method with 12 bacteria. The 3 active actinomycetes, selected from primary, secondary and cross streak method of antibacterial, anticandida activity were tested against thirteen fungi for antifungal activity by agar plug method and well diffusion method. Broad spectrum antimicrobial activity was confirmed by cross streak method for selected antagonistic actinomycetes.

Keywords: Antimicrobial activity, Screening methods, Mangroves, Rhizophora apiculata.

**DOI:** 10.20448/805.1.1.44.52

**Citation** | T. Janaki (2016). Antimicrobial Activity of Mangrove Actinomycetes from Soil Sample of Rhizophora Apiculata. Journal of Biotechnological Research, 1(1): 44-52.

Copyright: This work is licensed under a Creative Commons Attribution 3.0 License

Funding : This study received no specific financial support.

Competing Interests: The author declares that there are no conflict of interests regarding the publication of this paper.

History : Received: 9 June 2016/ Revised: 27 June 2016/ Accepted: 4 July 2016/ Published: 12 July 2016

Publisher: Online Science Publishing

#### **1. INTRODUCTION**

Actinomycetes are group of organisms that share both the characteristics of bacteria and fungi and they have high G+C content. They are the strongest antagonists among microbes. The antibiotic substances produced by them display antibacterial, antifungal, anticancer, antiprotozoic, antiviral, insecticidal properties. The antibiotics produced by the actinomycetes are safer than the antibiotics secreted by the fungi and bacteria. Bio active substances produced by the marine actinomycetes are reported by many researchers [5, ,17, 19, 26]. Few report that mangrove soil is a major source of actinomycetes [21, 23, 4, 7]. Mangrove

ecosystem is the most useful ecosystem diversified with variety of microbes. The search of new and novel antibiotics and other bioactive microbial metabolites is important for the fight against new emerging pathogens [1, 2]. Isolation of actinomycetes from unique unexplored natural habitats is of interest to avoid re-isolation of strains that produce known bioactive metabolites. Neglected habitats are proving to be a good source of novel actinomycetes and bio active compounds [9].

The present investigation aims at finding better antimicrobial compound for controlling the bacterial and fungal human diseases, with the help of mangrove actinomycetes that are selectively isolated from the soil near the root region of *Rhizophora apiculata*, from the Ariyankuppam back water area, Puducherry, India.

#### 2. MATERIALS AND METHODS

#### 2.1. Isolation of Mangrove Actinomycetes

Soil sample was collected near the root region of *Rhizophora apiculata*, from the Ariyankuppam back water area, Puducherry, India. Then the soil sample was air dried for 7-10 days at 40°C, crushed and sieved to remove the shells and debris and given to Soil testing laboratory, Department of Agriculture, Puducherry, for physico-chemical analysis. The soil sample was subjected to dryheat pretreatment (70°C for 15 min) [6, 7]. One gram soil was mixed and serially diluted in sterile water blanks. 0.1 ml of last dilution (10<sup>-6</sup>) was inoculated by pour plate method [27 using Starch Casein Agar [11] supplemented with Fluconazole 80µg/ml and Nalidixlic acid 75µg/ml. Plates were incubated at 30  $\pm$ °C for up to 30 days. Plates were regularly examined for actinomycetes colonies. Selected colonies were subcultured in YME agar slants.

#### 2.2. Screening of Actinomycetes for Antimicrobial Activity

#### 2.2.1. Test Organisms Used in This Study

The following test bacteria were procured from Microbial Type Culture Collection-Chandigarh. The gram negative bacteria are *Pseudomonas aeruginosa* (MTCC-424), *Shigella flexneri* (MTCC-1457), *Bordetella bronchiseptica* (MTCC-6837), *salmonella typhi* (MTCC-3220), *vibrio cholera* (MTCC-3906), *Proteus vulgaris* (MTCC-744), *E.coli* (MTCC-1687), *Klebsiella pneumonia* (MTCC-4031), *Pseudomonas fluorescens, Enterococcus faecalis* (MTCC-439) and gram positive bacteria are *Staphylococcus aureus* (MTCC-96), *Bacillus subtilis* (MTCC-441) and One unicellular fungi-*Candida albicans* (MTCC-183).

The multicellular fungi used were *Microsporum gypseum* (MTCC-4494), *Trichophyton mentagrophytes* (MTCC-8476), *Epidermophyton floccosum* (MTCC-7880), *Colletotrichum capsici* (MTCC-3414), *Aspergillus fumigatus* (MTCC-3377), *Aspergillus niger* (MTCC-872), *Fusarium oxysporum* (MTCC-1755) and *Rhizoctonia solani* (MTCC-1236), *Alternaria alternata, Aspergillus flavus, Aspergillus terreus, Curvularia lunata* and *Colletotrichum gloeosporioides*.

#### 2.3. Preparation of Test Organisms

Test bacteria were maintained in nutrient agar broth, pH-7. These were stored in refrigerator at 4°C for future use. 12-24 hours old bacterial liquid cultures and candida culture were used for antimicrobial study.

Test fungi were maintained in potato dextrose broth and in PDA slants, pH-7. These were stored in refrigerator at 4°C for future use. 3-5 days old fungal liquid cultures and plate cultures were used for antifungal study

#### 2.4. Invitro Screening for Antimicrobial Activity

Primary screening by agar plug method was studied by following Mohanraj et al, [14] Secondary screening by agar well diffusion method was done by using Murrey et al, [13] procedure and Cross streak method was studied by Lemos et al, [12].

#### 3. RESULTS AND DISCUSSION

#### 3.1. Isolation and Maintenance of Actinomycetes

	Table-1. Physico-chemical characteristics of the soil samples													
S. no	Soil sample		Parameters in physico-chemical characteristics											
		ph	Statu	Soil type		Available macro- nutrients (mg/g)			Available micro-nutrients (mg/g)					
			E.C (dsm- <sup>1</sup> )	Lime		N <sub>2</sub>	$P_2O_5$	K <sub>2</sub> 0	Cu	Zn	Mn	Fe		
1	Rhizophora apiculata	7. 5	0.5	N	CL	80 VL	0.77 VL	166 H	0.550 L	0.902 L	1.819 L	4.299 M		

N-Normal; CL-Clay Loamy; L-Low; VL-Very Low; M-Medium; H-High

The abundance of the organic matters, salinity and high degree of moisture content favour the prevalence of actinobacterial population and other life forms in the mangrove ecosystem, same was reported by Nag et al, [15]. The pH of mangrove soil sample collected from *Rhizophora apiculata* was 7.5, usually, the pH of the fresh soil samples was ranging from 7.5 to 8.2 is the optimum range of pH needed for actinomycetes to live and produce the antibiotic substances as secondary metabolites in the soil near to the root region of mangrove plants. The air drying of soil samples for 7-10 days at 50°C helped to eliminate the vegetative bacteria and fungi; it was coincided with the study of Williams et al, [25]. The soil analysis results showed that there were very low available Nitrogen and P<sub>2</sub>O<sub>5</sub>. Micro-nutrients like Zn, Cu and Mn were low; Fe was medium, K<sub>2</sub>O is high in their available form. This may due to the seasonal variations and fluctuating water levels enriching actinomycetes counts. Srivibool, [20] suggested that the abundance of actinomycetes in mangrove soil may actually be due to better quality of the sediments, described in terms of structure, pH and humic substances. Totally 22 actinomycetes were isolated from soil sample of *Rhizophora apiculata* by dry heat (70°C for 15 min) pretreatment method [7]. Dry heat method yielded bioactive actinomycetes for antimicrobial activity. The isolated actinomycetes grew well in yeast malt extract agar (ISP<sub>2</sub>) and some of the isolates produce soluble pigments.

#### 3.2. Antibacterial Activity of Actinomycetes from R.apiculata

The great majority of antibiotics that have been isolated in numerous screening programs concerned with the search for new therapeutic agents have been tested primarily for their activity against different bacteria [24]. Accordingly, ten gram negative bacteria and 2 gram positive bacteria, procured from MTCC, Chandigarh was used for antibacterial study. 22 actinomycetes were isolated from *R.apiculata* and that were subjected for primary screening against human pathogenic bacteria by agar plug method.

S.no	Isolate code			Measu	remer	t of z	one o	f inhik	oition i	n milli	meter		
		E.c	k.p	p.v	p.a	s.t	s.f	V.C	B.b	p.f	E.f	B.s	S.a
1	M46	-	-	-	-	-	-	-	22	-	-	2	-
2	M47	-	-	-	12	-	-	-	-	-	-	8	-
3	M48	-	-	-	-	-	-	8	-	-	-		-
4	M49	-	-	-	30	-	10	-	20	-	30	10	-
5	M50	-	-	-	20	8	-	12	20	-	-	20	-
6	M51	-	-	-	14	-	-	-	-	-	-	-	-
7	M52	8	-	-	-	6	-	-	-	-	-	10	6
8	M53	-	-	-	-	-	-	-	-	-	-	-	-
9	M54	-	-	-	-	4	-	-	14	-	-	-	-
10	M55	-	-	-	-	-	-	-	4	-	-	10	-
11	M56	-	-	-	-	-	8	-	-	-	-	-	-
12	M57	-	-	-	-	-	-	-	14	-	-	-	-
13	M58	-	-	-	-	-	-	-	20	-	-	6	6
14	M59	-	-	-	-	-	-	-	-	-	-	-	-
15	M60	-	-	-	10	-	-	-	10	-	-	-	-
16	M61	-	-	-	-	-	-	-	24	-	-	-	-
17	M62	-	-	-	-	-	-	-	26	-	-	6	-
18	M63	-	-	-	20	-	-	-	10	-	-	-	-
19	M64	-	-	-	-	-	-	-	9	-	-	-	-
20	M65	20			15		26		25		20	20	
21	M66	-	-	-	-	-	-	-	4	-	4	-	-
22	M67	-	-	-	-	-	-	-	8	-	-	-	-

Table-2. Primary screening of actinomycetes from R.apiculata against human pathogenic bacteria by agar plug method

Source: From Ph. D unpublished Thesis, T. Janaki- original research work

E.c-E. coli, K.p-Klebsiella pneumoniae, P.v-Proteus vulgaris, P.a-Pseudomonas aeruginosa, S.t-Salmonella typhi, S.f -Shigella flexneri, V.c-Vibrio cholera, B.b-Bordetella bronchiseptica, P.f-Pseudomonas fluorescens, E.f-Enterococcus faecalis, B.s-Bacillus subtilis, S.a-Staphylococcus aureus.

The total percentage of inhibition by actinomycetes against bacteria in primary screening was noted as *E.coli*-9%, *K. pneumoniae*-0%, *P.vulgaris*-0%, *P.aeruginosa*-31.8%, *S.typhi*-13.6%, *S.flexneri*-13.6%, *V.cholera*-9%, *B.bronchiseptica*-68.2%, *P.fluorescens*-0%, *E.faecalis*-13.6%, *B.subtilis*-40.9%, *S.aureus*-9%. Totally 20 (90.91%) actinomycetes showed antibacterial activity towards any one of the tested bacteria, 2 (9.09%) actinomycetes showed no antagonistic activity. From these, it was noted that the mangrove actinomycetes were strong in their antibacterial activity. Only 4 actinomycetes were selected from *R. apiculata* and that were subjected for secondary screening.

# 

#### Percentage of active and inactive mangrove actinomycetes

**Figure-1.** Percentage of active and inactive actinomycetes from *R.apiculata* **Source:** From Ph. D unpublished Thesis, T. Janaki- original research work

#### 3.3. Antibacterial Activity of Actinomycetes in Secondary Screening by Agar Well Diffusion Method

The isolates selected from primary screening were subjected for secondary screening by agar well diffusion method. The isolates which produced antibiotic compound in large quantity in liquid media were selected as potent isolates. Well diffusion method helped to study about the antibiosis from liquid media fast. It was noted that the antibiotic production and antibacterial potency of the actinomycetes in liquid media was varying from the antibiotic production and antibacterial potency in solid agar medium [10].

S.n	Mangrove									in mr				
0	plant	lsolates code	E.c	k.p	p.v	p.a	s.t	s.fl	V.C	B.b	p.f	E.f	B.s	S.a
1	Rhizophora	M49	12	8	10	38	-	16	10	-		36	18	8
2	apiculata	M50	-	-	-	-	-	-	10	-		22	12	8
3		M52	8	-	6	12	10	8	-	8	-	32	20	10
4		M65	12	6	12	25	12	20	12	18	-	32	16	10

Table-3. Antibacterial activity of active isolates in secondary screening by agar well diffusion method

Source: From Ph. D unpublished Thesis, T. Janaki- original research work

Four active isolates were selected for antibacterial activity in secondary screening by well diffusion method from 22 isolates of *R.apiculata*. The percentage of inhibition of tested bacteria by the actinomycetes were, *E.coli*-75%, *K. pneumoniae-50%*, *P.vulgaris*-75%, *P.aeruginosa*-75%, *S.typhi*-50%, *S.flexneri*-75%, *V.cholera*-75%, *B.bronchiseptica*-50%, *P.fluorescens*-0%, *E.faecalis*-100%, *B.subtilis*-100%, *S.aureus*-100%. It was found that *E.faecalis*, *B.subtilis*, *S.aureus* are totally controlled by all the four active actinomycetes (M49, M50, M52 and M65) in the secondary screening.

It was observed that some actinomycetes grew well in solid medium and had shown antagonistic activity effectively towards the tested bacteria by agar plug method, but same isolates in liquid medium did not show any antagonistic activity towards the tested bacteria by agar well diffusion method in secondary screening. In few cases of actinomycetes, number of inhibition of tested bacteria by the actinomycetes by agar plug method in primary screening coincided with agar well diffusion method in secondary screening. The isolates, strong in

their antibacterial activity both in agar plug method (Solid media) and in agar well diffusion method (Liquid media) were selected for confirmatory test for antibacterial activity.

#### 3.4. Anticandida Activity of the Actinomycetes

It was an effort taken to screen the mangrove actinomycetes for anticandida activity to find better alternative. Out of 22 isolates from *R.apiculata*, 8 (36.36%) isolates were active and 14 (63.64%) were inactive for anticandida activity by agar plug method in the preliminary screening.

S.no	Isolate code	Inhibition in millimeter
1	M46	-
2	M47	-
3	M48	10
4	M49	24
5	M50	-
6	M51	-
7	M52	12
8	M53	-
9	M54	-
10	M55	-
11	M56	12
12	M57	-
13	M58	-
14	M59	10
15	M60	-
16	M61	-
17	M62	8
18	M63	
19	M64	-
20	M65	22
21	M66	-
22	M67	12

Table-4.	Anticandida	activity of	the actinomycetes
----------	-------------	-------------	-------------------

Source: From Ph. D unpublished Thesis, T. Janaki- original research work

#### 3.5. Secondary Screening of Actinomycetes for Anticandida Activity

8 isolates from *R.apiculata* with anticandida activity were selected for secondary screening to test their ability to produce the active compounds in liquid medium. It was witnessed that some isolates were not produce active compound for anticandida activity in liquid medium and that type of cultures were omitted after the secondary screening. The isolates that produced active compound both in solid and in liquid medium were taken for the final study as the active isolates for anticandida activity.

S.no	Mangrove plants	Isolates code	Zone of inhibition in mm
1	Rhizophora apiculata	M49	22
2		M52	14
3		M65	20

Source: From Ph. D unpublished Thesis, T. Janaki- original research work

Based on the secondary screening, 3 isolates from *R.apiculata* with strong activity were selected and subjected to confirmatory screening using cross streak method. The output of our study related to anticandida activity is better than the study of Susithra et al, [22]. Our study report that the actinomycete isolates efficiently controlled the *Candida albicans* under in-vitro conditions. This output confirmed that the mangrove sources are precious sources for anticandida drug delivary from actinomycetes.

#### 3.6. Antifungal Activity of Selected Isolates

The 3 active actinomycetes, selected from antibacterial, anticandida activity were tested against thirteen fungi for antifungal activity by agar plug method and well diffusion method. Results indicated that the mangrove actinomycetes are very active in controlling the growth of both phytofungal pathogens and zoophilic fungal pathogens [9]. Actinomycete-fungus antagonism has been demonstrated for a wide variety of plant pathogens such as *Alternaria sp.* Chattopadhyay and Nandi, [3] *Rhizoctonia sp.* [18] and *Cuvvularia sp.* [16].

Isolate	Fungi used in antifungal activity, Inhibition in mm												
code	M.g	T.m	E.f	C.I	A.a	A.fu	A.n	A.f	A.t	R.s	C.ca	C.g	F.o
M49	8	4	6	12	8	-	-	-	-	-	10	8	6
M52	-	-	-	-	-	-	6	4	8	-	-	-	-
M65	4	6	4	-	-	-	12	-	-	12	8	14	12

Table-6. Antifungal activity of selected isolates by agar plug method

M.g- Microsporum gypseum, T.M- Trichophyton mentagrophytes, E.f- Epidermophyton floccossum, C.I- Curvularia lunata, A.a- Alternaria alternata, A.fu-Aspergillus fumigatus, A.n- Aspergillus niger, A.f- Aspergillus flavus, A.t-Aspergillus terrus, R.s-Rhizoctonia solani, C.ca-Colletotrichum capsici, C.g-Colletotrichum gleosporioides, F.o-Fusarium oxysporum

#### 3.7. Cross Streak Method to Confirm the Antimicrobial Activity of Selected Isolates

Cross streak method was used for confirmation of antimicrobial activity for most active isolates. The antibacterial and anticandida activity of the isolates was studied better in nutrient agar plates and antifungal activity was studied with PDA plates for cross streak method. It was observed that inhibition of bacteria by isolates in cross streak method was better than the agar plug method, because the antibiotic compound in the cross streak plates was not disturbed, the whole production of antibiotic compound was in the same plate but in the agar plug method only 8mm radial agar plugs were cut and tested for antibacterial activity [10]. The antimicrobial activity of actinomycetes was varying from one method to other in antimicrobial screening.

#### 4. CONCLUSION

Mangrove actinomycetes from *Rhizophora mucronata* inhibited both gram positive as well as gram negative bacteria efficiently and able to lyse and destroy the cell wall types of both gram +ve and gram –ve bacteria. The gram–ve bacteria; *Pseudomonas aeruginosa, Bordetella bronchiseptica* and gram positive bacteria; *Bacillus subtilis* were more sensitive to the mangrove actinomycetes from *Rhizophora mucronata*. The overall sensitivity of bacteria to isolates added valuable information that the mangrove soil is the efficient source for isolating potent antinomycete isolates for the bacteria those involve in causing nosocomial infections in human beings. Anticandida study revealed information that the actinomycete isolates from

mangrove sources are very effective and it can be used in the pharmaceutical field for anticandida drug delivary. Antifungal activity added valuable information about mangrove actinomycetes and its application in the field plant protection and pharmaceutical field for human welfare.

#### REFERENCES

- [1] Berdy, J. (2005). Bioactive microbial metabolites, J. Antibiot., 58(1): 1-26.
- [2] Busti, E., Monciardini, P., Cavaletti, L., Bamonte, R., Lazzarini, A., Sosio, M. and Donadio, S. (2006). Antibiotic producing ability by representatives of a newly discovered lineage of actinomycetes. Microbiol., 152: 675-683.
- [3] Chattopadhyay, S.K., and Nandi. (1982). Inhibition of Helminthosporium oryzae and Alternuria by Strptomyces longisporus (Krassilnikov) Waksman. Plant Soil. 69: 171-175.
- [4] Dhanasekaran, D., Panneerselvam, A. and Thajuddin, N. (2008). An antifungal compound: 4' phenyl-1-napthyl– phenyl acetamide from Streptomyces spp.DPTB16. Facta Universitatis Series: Medicine and Biology, 15: 7-12.
- [5] Farooq Biabani, M.A., Laatsch, D., Helmke, E., Weyland, H., (1997). Δ-Indomycinone: a new memberof pluramycin class of antibiotics isolated from marine streptomyces sp. J.Antibiot. 50: 874-877.
- [6] Hayakawa, M., Sadaka, T., Kayiura, T. and Nonomura, H. (1991). New methods for the highly selective isolation Micromonospora and Microbispora. Journal of Fermentation and Bioengineering, 72: 320-326.
- [7] Janaki .T Nayak B.K and Ganesan.T. (2014) Different Pre-treatment methods in Selective Isolation of Actinomycetes from Mangrove sediments of Ariyankuppam, Back water Estuary, Puducherry. Int. J. Adv. Res. Biol. Sci. 1 (6):154-163.
- [8] Janaki .T Nayak BK and Ganesan .T (2014). Antibacterial activity of Mangrove Actinomycetes isolated by Eight Different pre-treatment methods from backwater estuary, Ariyankuppam, Puducherry. Int.J.Pharm Res & Bio. 3(6): 132-149.
- [9] Janaki .T Nayak. BK and Ganesan. T. (2016). Antifungal activity of soil actinomycetes from the mangrove Avicennia marina. Journal of Medicinal Plants Studies. 4(2): 05-08.
- [10] Janaki.T Nayak. BK and Ganesan. T. (2016). Antibacterial activity of soil actinomycetes from the mangrove Avicennia marina. Journal of Pharmacognosy and Phytochemistry, 5(1): 267-271.
- [11] Kuster E. and Williams S.T. (1964). Selection of media for isolation of streptomycetes, Nature, 202: 928-929.
- [12] Lemos ML, Toranzo AE, Barja JL. (1985). Antibiotic activity of epiphytic bacteria isolated from intertidal seaweeds. Microbial Ecol , 11: 149-163.
- [13] Murrey, P.R., Baron, E.J., Pfaller, M.A., Tenover, F.C., Yolken, H.R., (1995). Manual of Clinical Microbiology, 6thEdition. ASM Press, Washington, DC, 15-18.
- [14] Mohanraj D., Bharathi S., Radhakrishnan M., Balagurunathan R. (2011). Bioprospecting of actinobacteria from Yelagiri hills with special reference to antibacterial activity. J. Chem. Pharm. Res. 3(3): 439-446.
- [15] Nag Chaitali, Bhattacharya Sourav and Das Arijit, (2012). Evaluation of antagonistic activities of microbes from Vallapattanam and Pappinishery mangrove ecosystems of Kannur district in Kerala, India. Int. J. of Pharm. & Life Sci. 3(5): 1650-1659.
- [16] Paul, A.K., and A.K. Banerjee. (1986). In vitro effect of an antifungal antibiotic produced by Streptomyces galbus
  5ME-13. Hind. Antibiot. Bullet. 28: 15-19.

- [17] Pusecker, K., Laatsch, H., Helmke, E., Weyland, H., (1997). Dihydropencomycin methyl ester, a new phenazine derivative from a marine streptomycete. J. Antibiot. 50: 47-483.
- [18] Rothrock, C.S and D. Gottlieb. (1984). Role of antibiosis in antagonism of Streptomyces hygroscopicus var geldanus to Rhizoctonia solani in soil. Can. J. Microbiol. 30: 1140-1 147.
- [19] Romero, F., Espliego, F., Baz, J.P., De Quesada, T.G., Gravalos, D., De La Calle, F., Fernandez-Puentes, J.L. (1997). Thiocoraline, a new depsipeptide with anti-tumour activity produced by a marine micromonospora. J. Antibiot.; 50: 734-737.
- [20] Srivibool, R. (2000). Antimicrobial activity of Actinomadura isolates from tropical island soils. Actinomycetes. 10:10-12.
- [21] Sivakumar, K., Sahu, M. and Kathiresan, K. (2005). Isolation and characterization of streptomycetes producing antibiotic from mangrove environment. Asian Journal of Microbial Biotechnology and Environmental Science; 7: 457-764.
- [22] Susithra M.P., Thenmozhi, M and Kannabiran K. (2009). Anticandidal activity of streptomyces paraguyensis isolated from marine sediment samples collected at the puducherry coast, Bay of Bengal, India. Pharmacologyonline 2: 527-537.
- [23] Vijayakumar, R., Muthukumar, C., Thajuddin, N., Pannerselvam A. and Saravanamuthu R. (2007). Studies on the diversity of Actinomycetes in the Palk Strait region of Bay of Bengal, India. Actinomycetologica, 21: 59-65.
- [24] Waksman, S. A., Lechevalier, H.A. Romano, A.H and Raubitschek, F. (1952). Antifungal antibiotics. Bull.org. mond.sante. Bull.wld. hth. Org. 6: 163-172
- [25] Williams, ST., Shameemullah, M., Watson, ET., Mayfield, CI. (1972). Studies on the ecology of actinomycetes in soil. VI. The influence of moisture tension on growth and survival. Soil Biol Biochem 4: 215–225
- [26] Williams, D.E., Bernan, V.S., Ritacco, F.V., Maiese, W.M., (1999). Holyrines A and B, possible intermediates in staurosporine biosynthesis produced in culture by a marinr actinomycete obtained from the North Atlantic ocean. Tetrahedron Lett.; 40: 7171-7174.
- [27] Zheng, Z., W. Zeng., Y. Huang, Z. Yang, J. Li, H. Cai, W. Su. (2000). Detection of antitumor and antimicrobial activities in marine organism associated actinomycetes isolated from the Taiwan Strait, China. FEMS Microbiol. Lett. 188: 87-91.

**Online Science Publishing** is not responsible or answerable for any loss, damage or liability, etc. caused in relation to/arising out of the use of the content. Any queries should be directed to the corresponding author of the article.